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Silver fir (*Abies alba*) trunk extract protects guinea pig arteries from impaired functional responses and morphology due to an atherogenic diet

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A B S T R A C T

Background: Diet, rich in plant polyphenols prevents atherogenesis that manifests as reduced vascular relaxation and formation of plaques.

Hypothesis: Atherosclerosis could be reduced by the intake of silver fir (*Abies alba*) extract (SFTE), rich in polyphenols.

Study design: Chronic, *in vivo* treatment animal study.

Methods: Guinea pigs (*Cavia porcellus*) were fed for 8 weeks with one of the following three diets: atherogenic, basic or atherogenic + SFTE. After isolation, we measured the relaxation and contractile responses of the thoracic aorta. Additionally, we measured the area of fatty plaques on the aortic walls.

Results: Compared to the basic diet, the atherogenic diet decreased the ability of the aorta to relax by 63% (p < 0.001). The addition of SFTE to the atherogenic diet improved the aorta relaxation response compared to that of the atherogenic diet without SFTE (the decrease relative to the basic diet was $26\%, p < 0.001$). The aorta contractility did not differ between the groups. The SFTE group generated significantly fewer atherosclerotic plaques than did the atherogenic group. The areas of atherosclerotic plaques were 7.4, 0.3 and 1.6% in the aortas of guinea pigs receiving atherogenic, basic or atherogenic $+$ SFTE diets, respectively.

Conclusions: In a guinea pig model, prolonged treatment with antioxidative polyphenol-rich SFTE prevents aortic functional and morphological changes caused by an atherogenic diet.

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1 **Introduction**

Atherosclerosis is the major precursor of cardiovascular disease $\overline{2}$ and is a chronic inflammatory process in arterial walls that is caused by the accumulation of macrophages and low-density lipoproteins. The interaction between plasma cytokines, lipoproteins and artery- [specific proteins influences lesion initiation and growth \(Pelosi et al.](#page-4-0) 2014). Atherosclerosis is characterized by elevated total cholesterol

8 and low-density lipoprotein cholesterol. The condition is a chronic

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<http://dx.doi.org/10.1016/j.phymed.2015.06.004> 0944-7113/© 2015 Published by Elsevier GmbH. disease that can remain asymptomatic for decades and that can be 9 prevented by a healthy lifestyle. Strong evidence indicates that the 10 inflammation of the blood vessel intima is caused by reactive oxygen 11 species (ROS), which form upon oxidative stress. This response repre- 12 sents a state of imbalance between the production and elimination of 13 free radicals that, in excessive quantities, damage tissues. In addition 14 to pollution, smoking, exercise deficiency and stress, one of the ma- 15 jor causative factors of atherosclerosis is a Western-type diet rich in 16 saturated fats and poor in fiber and antioxidants [\(Miller et al. 2013\)](#page-4-0). 17

Antioxidants are compounds or enzymes that are capable of 18 counteracting the damaging effects of oxidation. Antioxidative and 19 anti-inflammatory plant phenols have been demonstrated to re- 20 duce atherosclerosis and improve endothelial function (Stoclet et al. 21 [2004\). Most studies have focused on catechins from green tea, co-](#page-4-0) 22 coa and maritime pine bark extract because they lower the intesti- 23 [nal absorption of lipids and decrease cholesterol synthesis \(Moore](#page-4-0) 24 et al. 2009; Salvamani et al. 2014). Polyphenols decrease oxidative 25 stress, prevent inflammation, and reduce platelet aggregation and the 26

Abbreviations: SFTE, silver fir trunk extract; ROS, reactive oxygen species; EDTA, ethylenediaminetetraacetic acid; TCA, trichloroacetic acid; TBA, thiobarbituric acid; LDL, low-density lipoprotein; NO, nitric oxide; NOS, nitric oxide synthase; (MCP)-1, monocyte chemoattractant protein-1; VLDL, very low-density lipoprotein; BHT, butylated hydroxytoluene; PBMC, peripheral blood mononuclear cell; DPPH, 2,2-diphenyl-1-picrylhydrazyl; CVD, cardiovascular disease.

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 [proliferation of vascular smooth muscle cells \(Curin and Andriantsi-](#page-4-0) tohaina 2005). Polyphenols also act as beneficial agents in cardiovas- cular disorders, diabetes mellitus, rheumatism, chronic venous insuf- ficiency and other inflammatory diseases (Enseleit et al. 2012; Gulati [2014; Maimoona et al. 2011\). Among other mechanisms, cardiovascu-](#page-4-0) [lar drugs also appear to act through scavenging effects \(Marton et al.](#page-4-0) 33 2001).

 Silver fir trunk extract (SFTE) contains a complex mixture of bioac- tive polyphenols from the trunk of the silver fir tree (*Abies alba*), of which the main constituents are catechins, phenolic acids and lig- nans. SFTE exhibits strong *ex vivo* antioxidative activity when in- cubated with primary human peripheral blood mononuclear cells 39 (Tavčar Benković et al. 2014); however, no scientific data have been published regarding SFTE's efficiency under physiological conditions. Other studies on *Abies alba* wood revealed the potency of ethyl ac- etate extract in scavenging free radicals and chelating ferrous ions [\(Vasincu et al. 2013\)](#page-5-0). An ideal antioxidant should be readily absorbed by the body and should prevent or quench free radical formation or [chelate redox metals at physiologically relevant levels \(Poljsak et al.](#page-4-0) 46 2013).

 As in humans, Western-type diets can induce atherosclerosis in certain rodent models. Among these models, guinea pigs exhibit a cholesterol profile that most closely resembles that of humans when challenged with a high-cholesterol diet. Therefore, guinea pig models [are valuable for testing the efficiency of various therapies \(Fernandez](#page-4-0) and Volek 2006).

53 In this study, we demonstrated that SFTE could prevent morpho-54 logical and functional changes of the arterial wall of guinea pig aortas 55 as a result of an 8-week atherogenic diet.

56 **Material and methods**

57 *Silver fir trunk extraction*

 Silver fir trunk extract, characterized in our previous studies was prepared by the following two-step process, according to a modifica-60 tion of a previously published procedure (Strukelj et al. 2012; Tavčar 61 Benković et al. 2014): 5 kg of the ground trunk of the silver fir (*Abies alba* Mill., checked with [www.theplantlist.org\)](http://www.theplantlist.org) was extracted with 25 l of water at 70°C for 2 h. The aqueous extract was then evapo- rated under vacuum to a volume of 5 l. In the second step, the con-65 centrated aqueous extract was extracted with 3×3 l of ethyl acetate. Twenty-five milliliters of polyethylene glycol 400 was added to the ethyl acetate extract, and the ethyl acetate was then evaporated from the mixture. We obtained 50 ml of viscous, liquid SFTE. According to EMA guideline [\(European Medicines Agency \(HMPC\) 2010\)](#page-4-0), the ex- tract is "other herbal preparation" declared as: refined liquid extract from *Abies alba* Mill., truncus (DER = 100 : 1). Extraction solvent: 72

 As recommended in EMA reflection paper (European Medicines [Agency \(HMPC\) 2008\), protocatehuic and p-coumaric acids were cho-](#page-4-0) sen as analytical markers since they are potentially connected to the biological activity of the extract and reference compounds are avail- able for their quantification. Their specific analysis was carried out 78 by a validated HPLC method (Tavčar Benković et al. 2014). The con- tent of protocatechuic acid was 7.7 g/l and the content of p-coumaric acid was 3.7 g/l. The extract was further characterized by a HPLC fin-gerprint chromatogram (Fig. 1).

82 *Determination of hydroxyl radical scavenging activity*

83 The ability of the extracts to inhibit nonsite-specific hydroxyl 84 radical-mediated peroxidation was carried out according to a previ-85 ously described method [\(Hinneburg et al. 2006\)](#page-4-0). The reaction mix-86 ture contained 200 μ l of extract dissolved in phosphate buffer (0.2 M, 87 pH 7.4), 200 μl of 1 mM FeCl₃ (dissolved in water), 100 μl of 1 mM

ascorbic acid solution in phosphate buffer, 100 μ l of 1 mM EDTA in 88 phosphate buffer, 200 μ l of 14 mM 2-deoxy-D-ribose in phosphate 89 buffer and 100 μ l of 10 mM H₂O₂ in phosphate buffer. Each tube was 90 vortexed and incubated at 37°C for 60 min, after which 1 ml of 10% 91 TCA with 0.5% TBA mixture was added. The samples were heated in 92 a water bath at 85°C for 30 min, and the extent of oxidation was es- 93 timated based on the absorbance of the solution at 535 nm. The per- 94 centage inhibition values were calculated using the absorbances of 95 the control (Ac) and sample (As); the controls contained all of the re- 96 action reagents except for the extract or positive-control substance. 97

Animal studies 98

The experiments were conducted in accordance with the guide- 99 lines of the Veterinary Administration of the Republic of Slovenia 100 (Permit No. 34401-23/2009/3), which conform to the Guide for the 101 Care and Use of Laboratory Animals from the Institute for Laboratory 102 Animal Research, National Research Council, Washington D.C. (Na- 103 tional Academy Press, 1996). 2014 104

Eighteen Dunkin Hartley guinea pigs (*Cavia porcellus* L.) of both 105 sexes, aged between five and eight months, were housed at a con- 106 stant ambient temperature ($24 \pm 1^{\circ}$ C) and under a regular 12:12 h cir- 107 cadian cycle. The male and female subiects were randomly assigned 108 cadian cycle. The male and female subjects were randomly assigned to one of three experimental groups, and 2–4 animals were kept in 109 each cage. Each animal had unlimited access to water and to one of 110 the following feeds: the atherogenic diet (2 males, 3 females) (38.5% 111 Altromin 3123 (Lage, Germany) guinea pig maintenance diet pellets, 112 38.5% Altromin 3113 (Lage, Germany) guinea pig breeding diet pellets, 113 8.6% yolk (Mercator, Slovenia), 5% lard (Mercator, Slovenia), 8.4% fruc- 114 tose (KEFO, Ljubljana, Slovenia), 1% cholesterol (Acros Organics, Bel- 115 gium)); the basic diet (3 males, 3 females) (100% Altromin 3123 (Lage, 116 Germany) guinea pig maintenance diet pellets); or the atherogenic 117 diet (3 male, 4 female) (0.02% SFTE, thoroughly mixed with feed). At 118 an average feed consumption of 50 g/kg of body weight, the extract 119 intake corresponded to 10 mg of SFTE per kg of body weight. 120

After 8 weeks, the animals were sacrificed using $CO₂$ after a prior 121 injection of 8500 I.U. of heparin per animal (Krka, Novo Mesto, Slove- 122 nia). The thoracic aortas were isolated, rinsed of blood, dissected, and 123 cleansed of fat and connective tissue. The tissue was cut transver- 124 sally into 8 cylindrical rings (5 mm in length), using caution to pre- 125 serve the endothelium. The aortic rings were immediately mounted 126 in standard organ baths filled with a K-H solution of the following 127 composition (in mM): NaCl 118.8, NaHCO₃ 23.8, KCl 4.7, MgSO₄ 1.2 128 (Merck, Darmstadt, Germany), $KH₂PO₄$ 1.2, CaCl₂ 2.5 and glucose 11 129 (Kemika, Zagreb, Croatia). The abdominal aorta was isolated, rinsed of 130 blood, dissected, cleansed of fat and connective tissue, and preserved 131 in 10% formalin in buffered saline for morphological examination. 132

Vascular reactivity studies 133

After mounting, the rings were equilibrated at 20 mN resting ten- 134 sion for 90 min, with periodic adjustment of the ring to the desired 135

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 level, and the K–H solution was exchanged every 15 min. The rings were then contracted with 60 mM of KCl until stable contraction was obtained, after approximately 30 min, and then rinsed with K– H solution for 10 min. The KCl-induced contraction and rinsing was repeated one time. Phenylephrine and acetylcholine (Sigma–Aldrich Chemie, Steinheim, Germany) were dissolved in distilled water. For the relaxation measurements, the rings were precontracted with 143 100 μ mol/lof phenylephrine for 5 min until the contraction reached a plateau, and then the rings were relaxed by incubation with five cu-145 mulative final concentrations of acetylcholine (10^{-8} - 10^{-4} mol/l) for 2 min intervals. The tension was recorded after each interval. Finally, the rings were equilibrated with the K–H solution for 10 min.

 For the contraction measurements, the aortic rings were sequen- tially contracted six times with phenylephrine (final concentrations 150 of 10^{-9} - 10^{-4} mol/l) for 2 min intervals, and the tension was recorded after each interval.

 The vascular responses were processed and recorded on a Dewetron acquisition system (Dewetron, Graz, Austria) after analogue-digital conversion (NI PCI-6013; National Instruments, Austin, TX, USA) on the hard disk of a personal computer using the DeweSoft 6.1 software (Dewetron, Trbovlje, Slovenia).

157 *Atherosclerotic plaque area measurement*

 The abdominal aortas were cut longitudinally and dyed for 15 min with Oil red O solution (210 mg Oil Red O (Sigma-Aldrich O-0625, St. Louis, ZDA), 60 ml of isopropanol (Merck, Darmstadt, Germany) and 40 ml of water). The tissues were then flushed with water, mounted between two glass plates and scanned. The red atherosclerotic area was identified manually on the computer screen as a percentage 164 of the total arterial wall inner surface area with the ImageJ (Image Processing and Analysis in Java, National Institute of Health, ZDA) software.

167 *Data analysis*

 The statistical analysis was performed using GraphPad Prism 5.0 (GraphPad Software, San Diego, CA, USA). The values are expressed 170 as the mean \pm SEM for the *n* observations, where n represents the number of animals and *m* represents the number of aortic rings used in each of the studied groups. The relaxation responses of the arterial rings are expressed as a percentage relative to the phenylephrine pre- contracted aortic rings. The isolated thoracic aorta results were fitted and plotted using sigmoidal concentration-response curves. A two- way analysis of variance (ANOVA) with a Bonferroni's post hoc test 177 was used to perform the intergroup comparisons. A value of $p < 0.05$ was considered significant.

179 **Results**

180 *Determination of hydroxyl radical scavenging activity*

181 The hydroxyl radical scavenging activity of SFTE (IC50 = 2.9 μ g/
182 ml) was better than that of resveratrol (IC50 = 5.8 μ g/ml). 182 ml) was better than that of resveratrol (IC50 = 5.8 μ g/ml), 183 Butylated hydroxytoluene (BHT) (IC50 = 9.2 μ g/ml), vitamin E 183 Butylated hydroxytoluene (BHT) (IC50 = 9.2 μ g/ml), vitamin E
184 (IC50 = 10.1 μ g/ml) or epigallocatechin gallate (IC50 = 18.3 μ g/ml). $(IC50 = 10.1 \mu g/ml)$ or epigallocatechin gallate $(IC50 = 18.3 \mu g/ml)$.

185 *Animal studies*

186 The average final body weight of the animals after the experiment 187 was 1049 g for the atherogenic diet group, 923 g for the basic diet 188 group and 923 g for the atherogenic $+$ SFTE diet group. The differ-189 ences were statistically significant ($p < 0.05$).

190 The endothelium-dependent relaxation of the aortic rings precon-191 tracted with phenylephrine was measured by vasorelaxation tests 192 with acetylcholine. The relaxation of the thoracic aorta rings from

Fig. 2. Relaxation of pre-contracted thoracic aorta rings with the cumulative addition of increasing concentrations of acetylcholine. Each point on the curve represents the mean ± SEM. Asterisks and crosses represent significant differences (two-way ANOVA with Bonferroni post-hoc test): xxx for $p < 0.001$, xx for $p < 0.01$, x for $p < 0.05$ when comparing an atherogenic diet to atherogenic diet + SFTE; ∗∗∗ for p < 0.001, ∗∗ for p < 0.01, [∗] for p < 0.05 when comparing an atherogenic diet + SFTE to a basic diet.

Fig. 3. Areas under the curves of relaxation, as a function of acetylcholine concentration. One-way ANOVA with Bonferroni post-hoc test: $* p < 0.05$ when comparing an atherogenic diet to an atherogenic diet + SFTE, ** $p < 0.01$ when comparing an atherogenic diet to a basic diet.

the three groups of animals strengthened with increasing concentra- 193 tions of acetylcholine (Fig. 2). The extent of relaxation was dramati- 194 cally reduced in the aortic rings of the animals fed an atherogenic diet 195 and was highly improved with the inclusion of SFTE in the diet. The 196 differences between the groups were significant at the three highest 197 acetylcholine concentrations. The contractions of the contract

Fig. 3. presents the areas under the relaxation curves from 199 Fig. 2. The thoracic aorta of the guinea pigs fed an atherogenic diet 200 for 8 weeks demonstrated a 63% decrease ($p < 0.001$) in the re- 201 laxation response, compared to the animals that were fed the basic 202 diet. The addition of SFTE to the atherogenic diet significantly im- 203 proved the relaxation response. The aortic relaxation of the athero- 204 genic diet + SFTE group decreased by 26% (p < 0.001) in comparison 205
to the basic diet group. to the basic diet group.

The ability of the aortas to contract did not differ significantly 207 between the three groups receiving different diets [\(Fig. 4\)](#page-3-0). We ob- 208 served that some contraction values increased above 100% for the 209 basic group. We postulated that the extent of the phenylephrine- 210 stimulated contraction was greater than that of the KCl-stimulated 211 contraction in the basic group. The extent of the aortic ring contrac- 212 tion was identical for the atherosclerotic diet $+$ SFTE group. 213

The inner surface of the abdominal aortas from guinea pigs that 214 received an atherogenic diet for 8 weeks exhibited drastically in- 215 creased areas of atherosclerotic plaques in comparison to the animals 216 fed a basic diet [\(Fig. 5\)](#page-3-0). The addition of SFTE to the atherogenic diet 217

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Fig. 5. The area of atherosclerotic plaques, shown as the percent of the total area of the inner surface of the arterial wall. One-way ANOVA with Bonferroni post-hoc test: ∗∗∗ p < 0.001. The atherosclerotic plaque areas were 7.4, 0.3 and 1.6% in the aortas of guinea pigs receiving the atherogenic, basic and atherogenic + SFTE diets, respectively. The group that received SFTE in addition to the atherosclerotic diet generated significantly fewer atherosclerotic plaques than did the group receiving the atherogenic feed alone (typical walls of the aortas are shown in Fig. 6).

218 significantly decreased the atherosclerotic process, compared to that 219 in the animals receiving an atherogenic diet without SFTE.

220 **Discussion**

 In this study, the prophylactic effects, i.e., cardiovascular protec- tive effects, of SFTE on the functional and morphologic changes in the aorta that arise from a chronic atherogenic diet were studied. The effects of three different diets on the contraction and relaxation responses of thoracic aortas and on the development of atheroscle- rotic plaques in the abdominal aortas of guinea pigs were tested. The guinea pigs were divided into three groups and fed for 8 weeks with an atherogenic diet, basic diet or atherogenic diet + SFTE (Fig. 6). **Q3**²²⁸

The guinea pigs were shown to be one of the best models for eval- 229 uating atherosclerotic damage (Janić et al. 2014). Similar to humans, 230 these animals store most of their blood cholesterol in LDL form, pos- 231 sess similar enzymes associated with cholesterol synthesis and are 232 highly sensitive to dietary lipid intake [\(Fernandez and Volek 2006\)](#page-4-0). 233 A high-cholesterol diet successfully induced experimental athero- 234 genesis and caused significant changes in the guinea pig aortas af- 235 ter 8 weeks, with an average of 50 g of feed consumed per animal 236 per day. 237

We demonstrated that prolonged simultaneous feeding of an 238 atherogenic $+$ SFTE diet, which is rich in antioxidative polyphenols, 239 prevents the functional and morphological changes caused by an 240 prevents the functional and morphological changes caused by an atherogenic diet in the aortas of guinea pigs. Based on the litera- 241 ture, the observed effect can be achieved by one or more of the 242 following mechanisms: nonspecific effect of caloric restriction, in- 243 fluence on smooth muscle function (directly or via NO), influence 244 on the inflammation process (directly on macrophages or via cy- 245 tokines) and prevention of lipoprotein formation and oxidation. Each 246 of these possible mechanisms will be discussed in the following 247 chapters. 248

On average, the animals in all three groups consumed equal 249 amounts of feed (approximately 50 g per animal per day), but their 250 final body weights were significantly different. The reduced body 251 weight might have contributed to the observed prevention of func- 252 tional and morphological changes. 253

The functionality of the aortic rings depends on their amount of 254 smooth muscle, endothelial functionality and size, and pathological 255 changes in these vessels lower their functional responses. The SFTE- 256 rich diet significantly improved the relaxation response of the aortas. 257 In our previous study, SFTE was shown to increase the relaxation abil- 258 ity of rat aorta via a NO-dependent mechanism, as this effect was pre- 259 vented by a NOS inhibitor [\(Drevenšek et al. 2015\)](#page-4-0). Similar results were 260 obtained in studies using polyphenol-rich extracts from other plant 261 species, which improved endothelial function through increased NO 262 synthesis [\(de Pascual-Teresa et al. 2010; Maimoona et al. 2011\)](#page-4-0). The 263 phenols from strawberry leaves, kaempferol and rutin, were shown to 264 be direct, endothelium-dependent vasodilators that were mediated 265 by NO and cyclooxygenase products [\(Mudnic et al. 2009\)](#page-4-0). 266

In contrast, neither an atherogenic diet nor SFTE-rich diet influ- 267 enced the phenylephrine-initiated contraction of the aortas. Phenyle- 268 phrine is an α_1 agonist that induces the vasoconstriction of smooth 269 muscle cells through mechanisms in the endothelium and vascular 270 smooth muscle cells. We postulate that SFTE does not directly affect 271 smooth muscle cells; however, it can act through several other mech- 272 anisms. 273

We observed that SFTE reduced atherosclerotic plaque formation 274 by 80%. In previous studies, various plant polyphenols reduced ves- 275 sel wall inflammation and prevented the formation of atheroscle- 276 rotic plaques through different pathways (Leifert and Abeywardena 277 [2008\). In general, they reduced cholesterol absorption and accu-](#page-4-0) 278 mulation in macrophages [\(Curin and Andriantsitohaina 2005\)](#page-4-0). For 279

Fig. 6. Typical views of open abdominal aortas with visible atherosclerotic plaque areas from animals on the different diets. 1. Atherogenic 2. Basic 3. Atherogenic + SFTE.

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 example, a mixture of vitamins and minerals lowered LDL plasma levels, macrophage-positive areas and plaques in rabbits (Leborgne et al. 2005). In mice, quercetin inhibited monocyte chemoattrac- tant protein (MCP)-1, a critical contributor to the initiation and de- velopment of atherosclerotic lesions that directly promotes the mi- gration of inflammatory cells to the vascular wall. Quercetin also decreased plasma levels of interleukin 17, which acts as a potent mediator by increasing chemokine production in various tissues to recruit monocytes and neutrophils to the site of inflammation (Garelnabi et al. 2014). In ovariectomized guinea-pigs, polyphenol- rich grape intake altered hepatic cholesterol metabolism by lowering plasma triglycerides and VLDL cholesterol [\(Zern et al. 2003\)](#page-5-0). Cate- chin supplementation of apoE-deficient mice also reduced the mean atherosclerotic lesion area. These authors confirmed the atheropro- tective effect of catechin through the down-regulation of certain genes involved in energy metabolism, lipid metabolism and lipid traf- ficking (Auclair et al. 2009). Polyphenols also regulate various mech-anisms involved in inflammation [\(Tangney and Rasmussen 2013\)](#page-5-0).

 One of the explanations for SFTE efficiency is its strong an- tioxidant activity. *In vitro* tests revealed that SFTE is a better hy- droxyl radical scavenger than resveratrol, BHT, vitamin E or epigal- locatechin gallate. Our previous study evaluated the high ability of 302 SFTE to scavenge free radicals in PBMC cells (Tavčar Benković et al. [2014\), and those findings were supported by a study by Vasincu](#page-5-0) et al., who showed that ethyl acetate bark extract effectively scav- enged free radicals (DPPH, superoxide anions and hydroxyl radicals) and chelated ferrous ions [\(Vasincu et al. 2013\)](#page-5-0). In general, antioxi- dants reduce the formation of free radicals and other reactive oxy- gen species that can directly injure the arterial endothelium and cause lipid peroxidation of LDL particles, which then enter the blood vessel wall.

 Western-type diets are typical for some developed countries. High intake of saturated fats, sugar and low intake of fibers and antioxi- dants lead to elevated incidences of disease such as cardiovascular diseases (CVDs), diabetes, obesity and cancer (Everitt et al. 2006). In guinea pigs, we demonstrated that SFTE extract could prevent the damage caused by such diet. In accordance, many epidemiological and experimental studies have shown the beneficial role of antiox- idants in the diets of humans as prophylaxis against cardiovascular disease (Bertelli and Das 2009).

 After the strong *in vitro* potential for individual antioxidants such 321 as α -tocopherol, β -carotene and ascorbic acid was demonstrated, many clinical trials have reported disappointing results because an excessive number of single agents exhibited pro-oxidative activity (Lonn 2001). Our results indicate that complex, antioxidant-rich, nat- ural extracts possess several pharmacological activities and potential therapeutic advantages and might provide numerous benefits for hu-man health.

328 **Conclusion**

 SFTE added to an atherogenic diet of guinea pigs improved the re- laxation response and prevented the development of atherosclerotic plaques in aortas. The treatment did not influence aortic contractil- ity. SFTE was shown to be a potential agent in the prophylaxis and treatment of atherosclerotic changes.

334 **Conflict of interest**

 We wish to draw the attention of the editor to the following facts which may be considered as potential conflicts of interest. We re- ceived no financial contributions to this work. However, in the past 3 years, our department received financial support from following companies: Krka, d.d., Novo mesto; Lek d.d.; Medis, d.o.o.; ABIES LABS

340 razvoj in proizvodnja d.o.o.

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